Table I. HPLC Assay of Enantiometric Purity

cmpd	method	$t_{\rm R}$, min	ee/de, $%$	
L-Ser		17	98	
D-Ser		13	98	
		20	93	
e_{Dl} -8			95	

^a Daicel Chiralpak WH column at 50 °C eluting with 0.25 mM aqueous CuSO₄ at 2.0 mL/min and employing UV detection at 254 nm. b 10 μ m SiO₂ (25 cm × 4.5 mm) at 25 °C eluting with (20:1) hexanes-EtOAc at 2.0 mL/min and employing UV detection at 254 nm.

complished via flash chromatography on silica gel eluting with (3:2) hexanes-EtOAc and gave 90 mg (71% yield) of oxazolidine alcohol 7 as a colorless oil, $[\alpha]_D - 24.0^{\circ}$ (c 1.61, CHCl₃). An essentially identical procedure was applied to ent-5 and gave the antipode ent-7, $[\alpha]_D$ +23.6° (c 1.44, CHCl₃), in 64% yield: IR (neat) 3400, 1690, 1665 cm⁻¹; ¹H NMR (C₆D₆ + D₂O, 60 °C) δ 1.37 (s, 9 H), 1.43 (br s, 3 H), 1.56 (br s, 3 H), 3.51 (m, H), 3.62 (m, 3 H), 3.586 (m, H). Upon cold storage, a sample of ent-7 crystallized as prisms, mp 38-39 °C. Anal. Calcd for $C_{11}H_{21}NO_4$. C, 57.11; H, 9.17; N, 6.06. Found: C, 56.96; H, 9.21; N, 6.10.

B. Preparation of (-)-MPTA Esters of 7 and ent-7. To a solution of alcohol 7 (59 mg, 0.26 mmol), DCC (60 mg, 0.28 mmol), and DMAP (3 mg, 0.03 mmol) in dry CH_2Cl_2 (1.0 mL) was added 0.77 mL of a 0.38 M stock solution of (-)-MTPA in CH_2Cl_2 . After the mixture was stirred ambient temperature for

4.5 h, the TLC in (3:2) hexanes-EtOAc showed the clean formation of product 8, R_f 0.75 (UV and char B), at the expense of starting material at R_t 0.43. The resulting white suspension was filtered to remove the N , N' -dicyclohexylurea and then partitioned between EtOAc (20 mL) and $H₂O$ (10 mL). The organic layer was washed with 10 mL each of 1 \bar{N} HCl, H₂O, saturated NaHCO₃ solution, and brine then dried with $MgSO₄$, filtered, and concentrated to give 129 mg of crude product as an oily solid. Flash chromatography on silica gel eluting with (1:1) hexanes-EtOAc yielded 119 mg (104% crude yield) of material, $[\alpha]_D - 48^\circ$ (c 1.87, CHCl₃), that was analyzed directly: IR (neat) 1760, 1705 cm⁻¹; ¹H NMR $(C_6D_6, 75^{\circ}C)$ δ 1.39 (s, 9 H), 1.41 (br s, 3 H), 1.53 (br s, 3 H), 3.39 $(d, J = 1$ Hz, 3 H), 3.54 (dd, $J = 9.2$ and 5.8 Hz, H), 3.62 (dd, J = 9.2 and 1.9 Hz, H), 3.95 (m, H), 4.17 (m, H), 4.54 (dd, $J = 10.4$ and 3.2 Hz, H), 7.0-7.4 (m, 4H), 7.61 (br d, $J = 7.9$ Hz, H); ¹⁹F NMR (CDCl₃, 20 °C) δ 4.92 (s). An essentially identical procedure was performed with ent-7 and resulted in the isolation of epi-8 (see Table I), $\alpha|_D = 9.7^{\circ}$ (c 1.06, CHCl₃): IR (neat) 1750, 1700 cm⁻¹; ¹H NMR ($\ddot{C_6}D_6$, 75 °C) δ 1.41 (br s, 12 H), 1.55 (br s, 3 H), 3.38 (s, 3 H), 3.49 (dd, $J = 9.3$ and 5.6 Hz, H), 3.62 (dd, $J = 9.3$ and 1.9 Hz, H), 3.91 (m, H), 4.10 (m, H), 4.58 (dd, $J = 10.3$ and 3.4 Hz, H), 7.0–7.4 (m, 4 H), 7.60 (br d, $J = 7.3$ Hz, H); ¹⁹F NMR (CDCl₃, 19 °C) δ 4.84 (s), 4.99 (s).

Acknowledgment. We thank Andrew Terris for his technical assistance. This investigation was supported by Public Health Service Research Grant GM35557 from the National Institute of General Medical Sciences.

Novel Preparation of N-Protected Amino Acid Active Esters Using 1.2.2.2-Tetrachloroethyl Carbonates

Mahmoud Jaouadi, Jean Martinez,* and Bertrand Castro

Centre CNRS-INSERM de Pharmacologie Endocrinologie, 34094 Montpellier, France

Gérard Barcelo, Gérard Sennyey, and Jean-Pierre Senet

SNPE, Centre de Recherches du Bouchet, 91710 Vert-le-Petit, France

Received October 24, 1986

1.2.2.2-Tetrachloroethyl chloroformate reacts with substituted phenols or N-hydroxy imides to yield crystalline and stable mixed aryl or oximido tetrachloroethyl carbonates. When allowed to react with an N-protected amino acid derivative, these compounds proved to be efficient for the syntheses of the corresponding active esters. A series of active esters including p-nitrophenol, trichlorophenol, pentafluorophenol, and N-hydroxysuccinimide derivatives were prepared by this new procedure.

Active esters of amino acid derivatives represent one of the most important classes of activation for peptide coupling.¹ In a preceding paper, we presented a new method for the preparation of these active esters,² using 2-propenyl aryl carbonates, which constituted an alternative to the classical DCC coupling of N-protected amino acids with phenols. However, this method gave moderate yields in isolated active esters and was somewhat limited by the relatively expensive cost of starting 2-propenyl chloroformate material.

Following our investigation toward the application of new chloroformates in peptide synthesis, we turned our

attention toward 1,2,2,2-tetrachloroethyl chloroformate. This chloroformate is readily prepared (even on an industrial scale) by the reaction of chloral with phosgene^{3a} and has been used recently for the synthesis of N-protected amino acids.^{3b,c} In contrast to isopropenyl chloroformate, it is not suitable for direct mixed anhydride preparation. This is probably due to the instability of the intermediate mixed anhydride. Moreover, the chloral which is released in the reaction gives unwanted byproducts with the amino component. However, the mixed aryl or oximido tetrachloroethyl carbonates can be obtained by reaction of the

⁽¹⁾ Bodanszky, M. In The Peptides, Analysis, Synthesis, Biology (1) Consumers, M. H. H. H. H. H. H. H. (1998), M. H. (1998), M. (1998), M. (1998), M. (2) Jaouadi, M.; Selve, C.; Dormoy, J. R.; Castro, B.; Martinez, J. Tetrahedron Lett. 1985, 26, 1721.

^{(3) (}a) Cagnon, G.; Piteau, M.; Senet, J. P.; Olofson, R. A.; Martz, J. T. Eur. Pat. Appl. 40 153; Chem. Abstr. 1982, 96, 142281y. (b) Barcelo, G.; Senet, J. P.; Sennyey, G. J. Org. Chem. 1985, 50, 3951. (c) Barcelo, G.; Senet, J. P.; Sennyey, G.; Bensoam, J.; Loffet, A. Synthesis 1986, 627.

Table I. Mixed Aryl and Oximido Tetrachloroethyl Carbonate Derivatives

	method.	bp, $^{\circ}$ C (torr),	crystn	¹ H NMR,	IR,	
carbonate	yield $(\%)$	or mp, °C	solvent	ppm	cm^{-1}	combustion anal., found (calcd)
CCLCHCIOCOONSu	A, 83	$104 - 106$	pet. ether	2.9(s)	1760	C 26.00, H 1.74, N 4.10, O 23.54, Cl 43.55
	B. 83	108	Et ₂ O	6.6(s)		(C 25.87, H 1.55, N 4.31, O 24.6, Cl 43.55)
$CClsCHClOCOO(2,4,6)$ Tcp	A, 64	71	pet. ether	6.73 (s)		1800 C 26.61, H 1.02, O 11.56, Cl 60.80 (C 26.54, H
				7.4 _(s)		0.74, O 11.78, Cl 60.93)
$CCl3CHClOCOO(2,4,5)$ Tcp	A, 92	$150 - 155(0.02)$		6.7 (s)		1795 C 26.67, H 0.90, O 11.95, Cl 60.64 (C 26.54, H
				7.4 _(s)		0.74, O 11.78, Cl 60.93)
				7.56 (s)		
CCLCHCIOCOOPcp	A, 98	120	ethyl acetate	6.7(s)		1800 C 22.4, H traces, Cl 67.15 (C 22.66, H 0.27, Cl 67.05)
CCI ₃ CHCIOCOOPfp	A, 91	80(0.05)		6.7 (s)		1800 C 27.6, H 0.47, Cl 35.8, F 24.56, (C 27.44, H 0.26,
						Cl 36.00, F 24.12)
CCl ₃ CHClOCOON _p	C, 75	69	pet. ether	6.7(s)		C 30.87, H 1.53, N 3.81, O 22.23, Cl 41.15 (C
				7.5(d)	1790	30.97, H 1.44, N 4.01, O 22.92, Cl 40.64)
				8.36 (d)		
CCl ₃ CHClOCOO(2,4)DNp	C, 66	$121 - 122$	pet. ether	6.73 (s)		C 27.49, H 1.04, N 6.98, O 27.99, Cl 35.95 (C
				8.6 (dd)	1795	27.44, H 1.02, N 7.0, O 28.43, Cl 36)
				9.06 (d)		
				7.66 (d)		
CCI ₃ CHClOCOOBt	B , 85	$145 - 147$	CH_2Cl_2	7.04 (s)	1770	C 31.38, H, 1.37, N 12.16, Cl 41.03 (C 31.30, H
				7.6 (dd)		1.45, N 12.17, Cl 41.16)
				7.8 (dd)	1790	
				8.0(d)		
				8.2(d)		

Table 11. (tert -Butyloxycarbony1)amino **Acid N-Succinimidyl Esters**

above chloroformate with phenols **or** N-hydroxy imides according to reaction 1. These derivatives are able to **CIsCCHCIOCOCI** + **ROH** - **CIgCCHCIOCOOR (1)**

 \mathbf{a}

$$
R = -N
$$
. C₆C₁₅, C₆F₅, 2.4.5-C₆H₂C₁₃, 2.4.6-C₆H₂C₁₃,
4-C₆H₄NO₂, 2.4-C₆H₃(NO₂)₂, -N¹₁

produce active ester derivatives in the presence of a carboxylate (reaction 2). They have been found to be much

 $RCOOH + CI₃$ CCHCIOCOOR \rightarrow **RCOOR** + $CO₂$ + $CI₃$ CCHO (2)

$$
R = -N
$$
, C₆CI₅, C₆F₅, 2.4.5-C₆H₂CI₃

more suitable than the previously described aryl 2-propenyl carbonates for active ester synthesis of amino acid derivatives.

The mixed carbonate preparation has been performed by using 2,4,5- and 2,4,6-trichlorophenols, pentachlorophenol, pentafluorophenol, p-nitrophenol, 2,4-dinitrophenol, N-hydroxysuccinimide, and N-hydroxybenzotriazole. Yields are generally good and allowed large-scale preparations. The reaction with N-hydroxyphthalimide gave a mixture of the expected product together with 10% of diphthalimido carbonate. The reaction of 3-hydroxy-**4-oxo-3,4-dihydro-l,2,3-benzotriazine** gave an untractable mixture. Their physical properties are reported in Table I. The carbonates are usually crystalline and stable and can be stored at room temperature.

Some of these carbonates readily react with protected amino acid derivatives to yield the corresponding active esters, $CO₂$ and chloral. As chloral is quantitatively transformed to the water-soluble hydrate on hydrolysis, this new method provides a very clean, simple, and efficient method for active ester preparation. Compared to the classical preparation using DCC, elimination of the DCU filtration is an actual improvement. The results of the syntheses of various active ester derivatives are reported in Tables **11-v.** The best results were obtained with N-oxysuccinimidyl, 2,4,5-trichlorophenyl, and pentafluorophenyl derivatives; pnitrophenyl and pentachlorophenyl derivatives react sluggishly. Melting points and optical rotation were in good agreement with the lit-

Table **111. (Benzyloxycarbony1)amino** Acid N-Succinimidyl Esters

erature data. However, some pentafluorophenyl **esters** give significant deviations from the literature values. In our hands, reproduction of the described preparation using **DCC** confirms our data, but we cannot ascertain the absence of racemization in these cases. We assume however that these deviations are due to the pentafluorophenyl esters themselves rather than to the reagent. **As** described in the literature, 2,4-dinitrophenyl esters and N-oxybenzotriazolyl esters are very reactive and not very stable.

The reaction proceeds by initial attack of the **carboxylate** on the central carbonyl and release of either chloral and chloride ion or N-oxysuccinimide (or phenate) anion (Scheme I, steps a1 or bl). We have observed that when morpholine was allowed to react with 1,2,2,2-tetrachloroethyl N-succinimidyl carbonate, **1,2,2,2-tetrachloroethyl** N-morpholinecarboxylate and N-succinimidyl morpholinecarboxylate were formed in an 89/11 ratio (Scheme 11). Although the reaction is quite different, we assume that N-oxysuccinimide should be the best leaving group and route bl-b2 is the preferred mechanism. However, the transient oxyanion liberated in bl *can* act as a catalyst for the completion of the reaction through the al-a2 route.

In conclusion, this new method provides an easy preparation of N-protected amino acid active ester derivatives using cheap reagents, in a reaction where the byproduct is water soluble and easily eliminated from the reaction mixture.

Experimental Section

Capillary melting points are reported uncorrected. Thin-layer chromatograms (TLC) were **performed** on **silica** gel plates (Merck).

a RNH = morpioline.

Optical rotations were measured with Perkin-Elmer 241 MC polarimeter. Elemental analyses were performed by "Le Centre de Microanalyse du CNRS", ENSCM, Montpellier and ICSN, Gif

N-Protected Amino Acid Active Esters

sur Yvette. ¹H NMR spectra (60 MHz) were recorded on a Varian EM 360 A or (360 MHz) on a Bruker 360 spectrometer, at 25 °C, using $CDCl₃$ as a solvent and tetramethylsilane as an internal standard. Abbreviations used were those recommended by the IUPAC-IUB Commission *(Eur. J.* Biochem. **1984, 138,** 9-37).

General Procedure for the Synthesis of Mixed Aryl or N-Hydroxyaryl Tetrachloroethyl Carbonates. Method A. 1,2,2,2-Tetrachloroethyl chloroformate (12.35 g, **0.05** mol) was added to a stirred solution of the hydroxy compound (0.05 mol) in dichloromethane (50 mL). The reaction mixture was cooled to 0 "C, and pyridine (4 g, **0.05** mol) was added dropwise. The solution was allowed to warm at room temperature and stirred for 3 h. After addition of dichloromethane (150 mL), the reaction mixture was washed with cold water $(3 \times 50 \text{ mL})$. The organic layer was dried over MgS04 and concentrated in vacuo, and the residual product was distilled or crystallized **as** described in Table I.

Method B. In this method, pyridine was replaced by triethylamine. The reaction mixture was allowed to react for 1 h at 0 °C and then 2 h at room temperature. The reaction mixture was then treated as described in method A.

Method C. The required phenol (0.1 mol) was added to a mechanically stirred solution of **1,2,2,2-tetrachloroethyl** chloroformate (27 g, 0.11 mol) in a mixture of benzene (150 mL) and light petroleum ether (150 mL). The suspension was cooled to 0 "C, and triethylamine (11 g, 0.11 mol) was added dropwise with vigorous stirring. The reaction mixture was allowed to warm to room temperature and stirring was continued for 4 h. After filtration on Celite, the solution was concentrated in vacuo and the crystalline residue was rinsed with light petroleum ether and dried (Table I).

N-(*tert* **-Butyloxycarbonyl)-L-alanine Pentachlorophenyl Ester. 1,2,2,2-Tetrachloroethyl** pentachlorophenyl carbonate (2.27 g, 5 mmol) was added to a solution of N-(tert-butyloxycarbonyl)-L-alanine (0.84 g, *5* mmol) and triethylamine (0.7 mL, 5 mmol) in,THF (15 mL). The reaction mixture was stirred at room temperature for 2 h. Ethyl acetate (20 mL) was then added, and the organic solution was washed with a cold 10% citric acid solution (10 mL), then with a saturated potassium bicarbonate solution (10 mL), and finally with water (10 mL). The organic solution was dried on magnesium sulfate and evaporated in vacuo. The solid residue was crystallized from ethyl acetate/petroleum ether (2.06 g, 94%): mp 170 °C (lit.^{6a} mp 166 °C); $[\alpha]^{25}$ _D -24.5°

-
- **(8)** Dahno, W.; Li, C. H. *Int. J. Pept. Protein Res.* **1971, 3, 81. (9)** Kovacs, J.; Mayers, G. L.; Johnson, R. H.; Cover, R. E.; Ghatak,
- U. R. J. *Org. Chem.* **1970,35, 1810.**
	-
- **(10)** Ruttenberg, M. A. *J. Am. Chem. SOC.* **1968,90,5558. (11)** Zahn, **H.;** Danho, W.; Gutte, B. *2. Naturforsch., Anorg. Chem., Org. Chem., Biochem., Biophys., Biol.* **1966,21, 763.**

(c 1.0, CHCl₃) (lit.^{6a} [α]²⁵_D -22.2° (c 5.1, CHCl₃)).

N-(**tert -Butyloxycarbonyl)-L-phenylalanine** *p* **-Nitro**phenyl Ester. 1,2,2,2-Tetrachloroethyl p-nitrophenyl carbonate (0.87 g, 2.5 mmol) was added to a solution of N -(tert-butyloxy**carbonyl)-L-phenylalanine** (0.73 g, 2.75 mmol) and N-methylmorpholine (0.3 mL, 2.5 mmol) in THF (6 mL). The reaction mixture was stirred at room temperature for 1 h. Ethyl acetate (15 mL) was then added, and the organic solution was washed with a cold 10% citric acid solution *(5* mL), then with a saturated potassium bicarbonate solution (5 mL), and finally with water (5mL). The organic solution was dried on magnesium sulfate and evaporated in vacuo. The solid residue was crystallized from 95% ethanol (0.6 g, 62%): mp 128 °C (lit.^{6b} mp 132 °C); [a] -20.9 ° (c 2.0, DMF) (lit.^{6b} [α]_D -21.0 ° (c 2.0, DMF)).

General Procedure for the Synthesis of Amino Acid Active Ester Derivatives. To a solution of the N-protected amino acid (0.10 mol) in tetrahydrofuran (120 mL) or acetonitrile, containing N-methylmorpholine (0.11 mol), was added the mixed aryl or imide **1,2,2,2-tetrachloroethyl** carbonate derivative (0.10 mol). The reaction mixture was stirred at room temperature for 2-5 h. Ethyl acetate (500 mL) was added and the solution thoroughly washed with a 10% cold citric acid solution (1 **X** 100 mL), cold water (1 **X** 100 mL), a cold saturated bicarbonate solution (1 **x** 100 mL), and cold water (2 **X** 100 mL), dried over MgSO₄, and concentrated in vacuo at $t < 40$ °C. The residue, treated with the appropriate solvents (as given in the literature), leads usually to crystalline N-protected amino acid active esters. Their purity was ascertained by TLC in different solvents as indicated in the literature. They were identified by elemental analysis and comparison of their physical properties with those reported by other authors. The physical data of the synthesized N-protected amino acid active ester derivatives are reported in Tables 11-V.

Reaction of 1,2,2,2-Tetrachloroethyl N-Succinimidyl Carbonate with Morpholine. A solution of 1,2,2,2-tetrachloroethyl N-succinimidyl carbonate (1.63 g, **5** mmol) in THF (3 mL) was slowly added to a solution of morpholine (0.87 g, **10** mmol) in 3 mL of THF. The reaction mixture was stirred for 1 h and washed successively with 1 N HCl, 10% NaHCO₃, and water. The solution was dried over magnesium sulfate, evaporated to dryness, and the resulting product (9.5 g) examined by NMR *⁶*6.8 **(1** H, OCHCl), 3.4-3.8 (8.6 H, CH, of morpholine), 2.7 (0.5 H, $CH₂$ of succinimide). This is in accord with an 89/11 ratio of respectively **1,2,2,2-tetrachloroethyl** N-morpholinecarboxylate and N-succinimidyl morpholinecarboxylate.

(12) Pless, J.; Boissonnas, R. A. *Helu. Chim. Acta* **1963, 46, 1609. (13)** Broadbent, M.; Morley, J. S.; Stone, B. E. *J. Chem. SOC. C* **1967, 2632.**

(14) Fletcher, G. **A.;** Jones, J. H. *Int. J. Pept. Protein Res.* **1972,4,347. (15)** Anderson, J. C.; Kenner, G. W.; MacLeod, J. K.; Sheppard, J. C.

(16) Bentley, P. H.; Gregory, H.; Laird, **A.** H.; Morley, J. S. *J. Chem. Tetrahedron, Suppl.* **1966,** *No.* **8, 39.** *SOC.* **1964,6130.**

(17) Bodanszky, M.; Natarajan, S.; Hahne, W.; Gardner, J. D. *J. Med. Chem.* **1977,20, 1047.**

(18) Kisfaludy, L.; Low, M.; Nyeki, 0.; Szirtes, R.; Schon, 1. *Justus Liebigs Ann. Chem.* **1973, 1973, 1421.**

⁽⁴⁾ Anderson, G. W.; Zimmerman, J. E.; Callahan, F. M. *J. Am. Chem. SOC.* **1964,86, 1839.**

⁽⁵⁾ Schnabel, **E.;** Herzog, H.; Hoffmann, P.; Klauke, E.; Ugi, I. *Jwtus Liebigs Ann. Chem.* **1968, 716, 175. (6)** (a) Johnson, B. J.; Trask, E. G. *J. Org. Chem.* **1968,33,4521.** (b)

Sandrin, E.; Boissonnas, R. A. *Helv. Chim. Acta* 1963, 46, 1637. (c) Wuensch, E. In *Houben-Weyl's Methoden der organischen Chemie*; Thieme: Stuttgart, 1974; Vol. 15, Part II. (7) Laufer, D. A.; Blout, E. R. J. Am. Chem.